

UNIVERSITÀ DEGLI STUDI DI TORINO

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Scientific	Scientific	Host	Type of	Start of	Language	
area	responsible	Department	activity	mobility		
Physiology	Emilio Carbone	Dipartimento di Scienza e Tecnologia del Farmaco	Basic Research	April 2017	English	
Type of	Senior (equal c	1000 more than 10 y	ars old)			
fellowship	1 month					
Title of the	Altered excitation/inhibition balance in hippocampal neurons induced by mutations					
research project	of L-type Ca^{2+} channels associated to forms of autism and mental retardation					
Description	Neuronal L-type calcium channels (Cav1.2 and Cav1.3) are largely expressed at the					
of the	somatic and perisomatic levels of neurons in many brain areas, including the					
research	hippocampus [1]. Both channels activate at relatively low resting potentials (-50 to -					
project	40 mV) and thus allow Ca^{2+} to enter the neurons during basal and sustained					
	neuronal activity. As cytosolic Ca ²⁺ regulates a large number of intracellular					
	modulatory pathways, both channels appear strategic to control neuronal survival					
	and maturation. Both channels, indeed, are critical for regulating the outgrowth of					
	neuritis and development of excitatory and inhibitory synapses (synaptogenesis).					
	Cav1.2 and Cav1.3 channels are also critical for the control of somatic and					
	perisonatic neuronal excitability. This derives from their high expression density					
	and coupling to Ca^{2+} -dependent potassium channels: BK ("big K") and SK ("small K") channels. In this way, $Caul 2$ and $Caul 2$ a					
	K^{-}) channels. In this way, Cav1.2 and Cav1.3 regulate the waveform of action					
	potentials (AP), the frequency of tonic firing, the timing of AP bursts and the onset of AP synchronisms in hippocampal microaircuits. Any pathological alteration of					
	of AP synchronisms in hippocampal microcircuits. Any pathological alteration of Cav1.2 and Cav1.3 "gating" properties induces profound changes to neuronal					
	signaling and brain area functions					
	Among the neurodegenerative diseases associated to Cav1.2 and Cav1.2 above				and Cav1 3 channels	
	Among the neurodegenerative diseases associated to Cav1.2 and Cav1.3 channels, of particular interest are the point mutations that slow-down the "inactivation gate" of both channels. A reduced rate of inactivation causes increased Ca ²⁺ entry in neurons with consequent altered degree of excitability firing synchronism and					
	excitation/inhibition balance. These mutations in humans are associated to two rare forms of the autistic spectrum disorder (ASD): the Timothy Syndrome (TS) [2] and					
	the Primary A	Aldosteronism S	Seizers and Neur	ronal Abnormali	ities (PASNA) [3]	
	Both are asso	ciated to a glyci	ine to arginine n	nutation at positi	(G406R) for	
	Cav1.2 (TS) a	and at position 4	407 (G407R) for	Cav1.3 (PASN	A).	
	Using intracellular patch-clamp recordings and extracellular microelectrode arrays					
	(64 MEA) mo	onitoring, in this	s project we will	study the excita	ability properties of	
	single neurons, the response of excitatory and inhibitory synapses and the					
	synchronous	versus asynchro	nous activity of	hippocampal m	icrocircuits in wild-	
	type (WT) an	d knock-in (KI)	mice with the n	nutation G406R	Cav1.2 or G407R	
	Cav1.3. Cultu	ired hippocamp	al networks will	be studied at di	fferent days in vitro	
	(from day 11 ^t	^h to 22^{nd}). The	ΓS KI mice will	be available from	m prof. Randy	



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	Rasmusson (Buffalo, USA) while the PASNA KI mice will be available from prof.			
	Joerg Striessnig (Innsbruck, Austria).			
	Specifically, we will test how the Cav1.2 and Cav1.3 mutations alter the expression and the physiological properties of ion channels that control the AP firing at the somatic and perisomatic level (Cav, Nav, BK, SK and Kv) [4, 5]. We will also test how the two mutations causing autism affect the day at which the asynchronous activity of neurons switches to a synchronous burst activity of the developing hippocampal networks [6] and how they modify the synaptic plasticity of pyramidal excitatory neurons and inhibitory interneurons [7].			
	References			
	1. Marschallinger, J., et al., Cell Calcium, 2015. 58(6): p. 606-16.			
	2. Splawski, I., et al., Cell, 2004. 119 (1): p. 19-31.			
	3. Scholl, U.I., et al., Nat Genet, 2013. 45 : p. 1050-4. 4. Marcantoni A et al. Journal of Neuroscience 2010 30 (2): p. 491-504			
	 Vandael, D.H.F., et al., Journal of Neuroscience, 2012. 32(46): p. 16145-16359. 			
	6. Gavello, D., et al., Plos One, 2012. 7 (7).			
	7. Baldelli, P., et al., Journal of Neuroscience, 2005. 25 (13): p. 3358-3368.			
	8. Crepel, V., et al., Neuron, 2007. 54 (1): p. 105-120. 9. Ben-Ari Y et al. Neuroscientist 2012 18 (5): p. 467-486			
Profile	Expert researcher on ion channels physiology, pharmacology and molecular biology.			
Description	Technical experience on AP recordings from in vitro neurons with patch-clamp.			
1	MEAs and Ca ²⁺ imaging			
Research	With the help of the senior researcher, we expect:			
objectives	1) to determine how the two point mutations remodel the expression of the main			
-	ion channels regulating neuronal excitability (Cav, Nav, BK, SK and Kv).			
	2) to establish how the increased Ca^{2+} entry associated to the mutations could affect			
	the activation of the Ca ²⁺ -dependent BK and SK channels that condition somatic			
	AP firing and burst rhythms that drive synaptic activity.			
	3) to highlight the origins of the neuronal network synchronism that occurs at the			
	14-18 th day <i>in vitro</i> . We expect also to clarify if the synchronism is linked to			
	Cav1.2 and Cav1.3 expression [8] and how much is conditioned by GABAergic			
	interneurons maturation in culture [9].			
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